

Secretory IgA in Saliva can be a Useful Stress Marker

Satoshi TSUJITA and Kanehisa MORIMOTO

Department of Hygiene and Preventive Medicine, Osaka University School of Medicine, Osaka

Abstract

To evaluate secretory immunoglobulin A (sIgA) in saliva as an immunological stress marker, we reviewed the literature on sIgA and its variation caused by psychosocial factors. Among the studies on the effect of academic stress on sIgA secretion, we could distinguish two kinds of stress effects: the immediate stress effect which increases sIgA secretion immediately after stress, and the delayed stress effect which decreases sIgA secretion several days after stress. On the basis of production and secretion mechanisms of sIgA, we also speculated on possible mechanisms that underlie the variations of sIgA caused by stress. Eventually, we concluded that sIgA in saliva can be a useful stress marker if we analyze the delayed stress effect on sIgA separately from the immediate stress effect on sIgA.

Key words: IgA, secretory, marker, stress, saliva

Introduction

Psychoneuroimmunological research has shown that psychosocial factors, including stress, social support, and emotion may affect susceptibility to infectious disease by influencing the immune system^{1,2}.

Because the immune system is a part of a complex and interactive network formed by the brain, neurotransmitters and neuropeptides, secretory glands, and various types of immune cells, no single measure of 'immune functioning' can fully express immune competence. However, for practical and ethical reasons, only a few immunological parameters can be measured in experimental research dealing with human subjects.

In several studies, secretory immunoglobulin A (sIgA) has been chosen as a measure of resistance to infectious disease, because it plays an important role in the defense mechanism of mucosal membranes.

Besides, psychometric instruments (questionnaires) for measuring stress have been developed. To complement these questionnaires, immunological stress markers would be valuable objective measures.

This article reviews the literature on sIgA and its variations caused by psychosocial factors, speculates on possible mechanisms that underlie the variations of sIgA, and evaluates sIgA, especially salivary IgA, as an immunological stress marker.

General aspects of sIgA

Secretory immunoglobulin A (sIgA) is found in various secretory fluids, including saliva, breast milk, and nasal, gastrointestinal, bronchial, and urogenital secretions at high levels of concentration³⁻⁵.

The secretory process and immunological functions of sIgA have been extensively studied³⁻¹².

Briefly, the secretory process of sIgA can be summarized as follows. At first, dimeric immunoglobulin A molecules, joined by a glycoprotein named J chain, are produced locally by IgA producing plasma cells in the lamina propria of mucosal membranes or in the connective tissue of glands⁶⁻⁹. Some of them diffuse through basement membranes to the basolateral surface of epithelial cells, where they are taken up by the epithelial cells with polymeric immunoglobulin receptors (poly-Ig receptor), then transcytosed to the apical surface of the epithelial cells, and released into secretory fluids in the form of sIgA⁶⁻¹². This secretory process is shown schematically in Fig.1.

The sIgA molecule is composed of a dimeric immunoglobulin A containing the J chain, and another glycoprotein, secretory component (SC), which is a residue of the poly-Ig receptor and binds covalently to the J chain. The SC stabilizes the sIgA molecule and protects it from degradation by bacterial and digestive enzymes in the secretory fluid environment³⁻¹².

For immunological functions, sIgA antibodies prevent bacteria from forming colonies on mucosal surfaces, kill them directly or activate complements or provide synergism with innate defense mechanisms, e.g. lacto-ferrin, lacto-peroxidase, etc. They also neutralize toxins and enzymes produced by bacteria³⁻⁹. In addition, sIgA antibodies neutralize pathogenic viruses so as to inhibit their penetration into epithelial cells. Moreover, it is recently reported that even low concentrations of sIgA, which can not prevent influenza A virus from penetrating into cells, inhibit

Received Oct. 9 1998/Accepted Dec. 24 1998

Reprint requests to: Satoshi TSUJITA,

Department of Hygiene and Preventive Medicine, Osaka University School of Medicine, 2-2 Yamada-oka, Suita, Osaka 565-0871, Japan.

TEL & FAX: +81(6)6879-3923.

their infectivity by damaging their activation process in the cell¹³. It is also reported that dimeric IgA, endocytosed by the poly-Ig receptor, can neutralize the Sendai virus which penetrates into epithelial cells¹⁴.

Besides, the sIgA antibodies prevent allergens and carcinogens¹⁵ from being absorbed through mucosal membranes into the body. Moreover, immune-complexes formed beneath the epithelial cells after antigen absorption are transported out and into secretions by the poly-Ig receptor dependent transport system (or SC dependent transport system)¹⁶.

From these sIgA functions, it may be expected that high proper sIgA secretion would prevent infections by various microorganisms in the mucosal membranes effectively. In fact, a review article concluded that the relatively high levels of sIgA in saliva is related to the lower incidence of upper respiratory tract illness, with the average effective size $r = 0.25$ ¹⁷. In addition, it is

recently reported that transient salivary IgA-deficiency, which may permit various allergens to penetrate through the mucosal membranes, in the first year of life is a risk factor for the subsequent development of bronchial hyper reactivity¹⁸. Moreover, in children with a history of recurrent colds and flu, not only elevated psychosocial stress was observed but also lower sIgA/albumin ratios in saliva were detected¹⁹.

Effect of psychosocial factors on sIgA

Various psychosocial factors, including academic examination²⁰⁻²⁷, daily hassles²⁸, negative mood²⁹, desirable and undesirable daily events³⁰, work demand³¹, and various relaxing factors³²⁻³⁶ were investigated as a possible sIgA modifier. Results of those studies are shown briefly in Tables 1 to 3. In all of those studies, salivary IgA was chosen as a sIgA, because it is not only

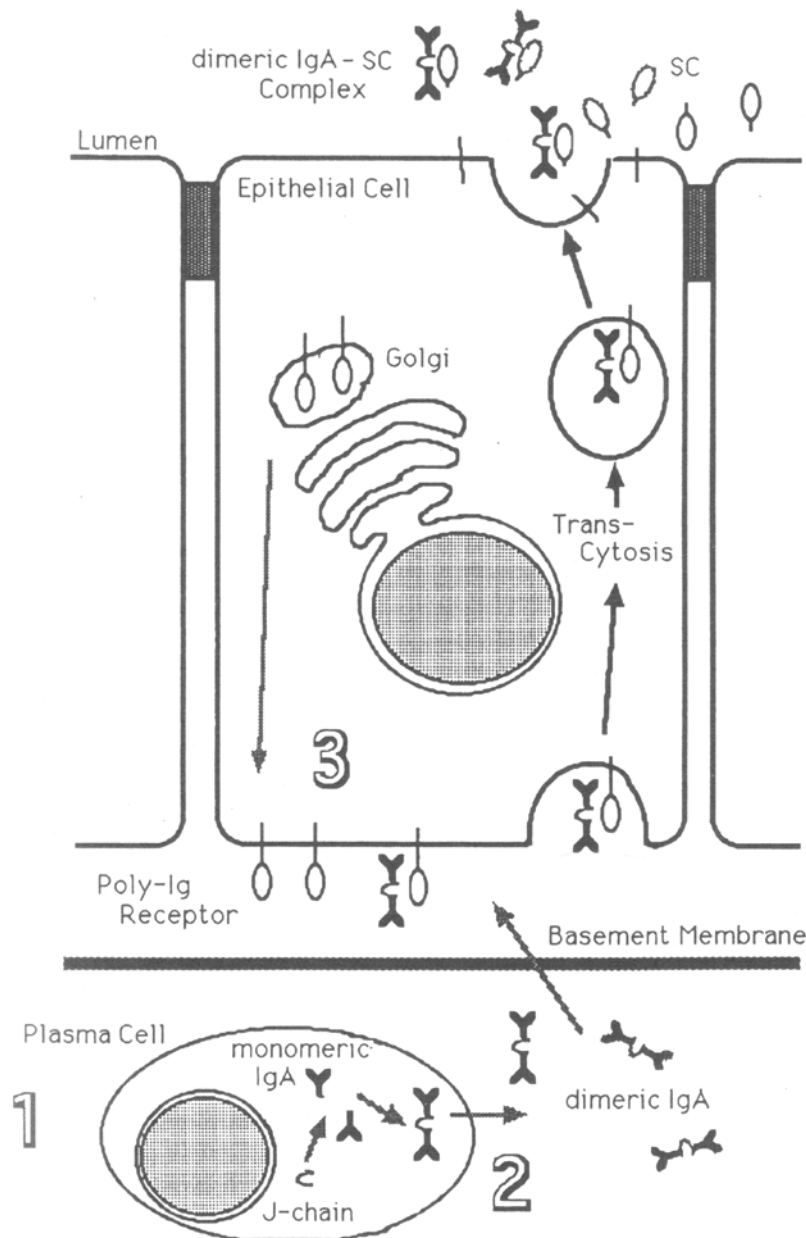


Fig.1 Selective transport system of IgA into external secretions. Psychosocial factors could affect sIgA response at the following three steps in the SC-dependent transport system: 1. number of plasma cells, 2. immunoglobulin producing activity of plasma cells, 3. expression of polyIg receptors or SCs.

important for defense against upper respiratory tract infectious diseases as mentioned previously, but it also can be collected easily.

However, current literature does not show much agreement about the determinant of a sIgA response, except for the relaxing factors. Studying academic stress, as shown in Table 1, a decrease in sIgA level²⁰⁻²³, as well as no alterations²⁴, or an increase²⁵⁻²⁷ have been reported. Studying other kinds of stressful events as shown in Table 2, the sIgA response was also not simple²⁸⁻³¹. On the other hand, all of the relaxing factors shown in Table 3 had positive effects on the sIgA response.

To explain the disagreement on the stress effect mentioned above, we tried to determine some rules which control the sIgA response to psychosocial factors. For simplifying the problem, we concentrated our attention on the studies on the academic stress effect as shown in Table 1. When we inspected the difference of methods of those studies, we noticed some interesting facts as follows. The sIgA levels decreased when it was determined several days or a few weeks after academic stress²¹⁻²³. Whereas the sIgA levels increased when determined immediately after academic stress²⁵⁻²⁷. (Refer to sIgA variation and sampling time in Table 1.) We may call those opposite sIgA responses after stress as a delayed

stress effect on the sIgA response and an immediate stress effect on sIgA response, respectively. The immediate stress effect seems to fade away in a short time after stress is relieved, resulting in a return of increased sIgA to its initial level^{25,27}.

As a result of this research, we have become able to understand, at least in part, the effect of other psychosocial stressors on sIgA secretion, as shown in Table 2. We may say it is a case of the delayed stress effect on the sIgA response that daily hassles caused a decrease in salivary IgA secretion after 4 weeks²⁸. We may also say it is a case of immediate stress effect on the sIgA response that the work demand of air traffic controllers caused an increase in salivary IgA secretion³¹.

However, it may not be appropriate to explain by the idea mentioned above that desirable and undesirable daily events correlate positively or negatively to salivary IgA secretion respectively³⁰, or that a negative mood showed a positive correlation to salivary IgA levels²⁹. We must have some other ideas to explain those emotional effects on salivary IgA secretion.

All of the relaxing factors shown in Table 3 had positive effect on the sIgA response³²⁻³⁶. We may call those phenomena a relaxation effect on sIgA response.

Table 1 Effects of academic stress on salivary IgA²⁰⁻²⁷.

Author	Stress	Change	Sampling time	Note
Jemmott (1983)	Exam	--(p<.025)	Five times over one year (Sept, Nov*, April*, June*, July)	Higher sIgA secretion in RAS group (p<.06). No recovery in IPS group.
Jemmott (1988)	Exam	--(p<.0001)	Three occasions (1st: 5 days before exam, 2nd*: on day of exam, 3rd: 14 days after exam.)	Higher sIgA concentration in adequate social support group (p<.05).
Mouton (1989)	Exam	--(p<.01) (at the most contrast, April-Sept)	Four occasions over 2 academic years (March, April*, Sept, Oct*)	A weak negative correlation (between stress level and sIgA secretion: r=-.25, stress level and sIgA concentration: r=-.36)
Deinzer (1998)	Exam	--(p<.01) weeks around exam	25 days before exam, and every day for 2 weeks around exam	Saliva was taken every morning immediately after awakening. No relationship between sIgA and URT symptoms.
Kiegolt-Glaser (1984)	Exam	+-	Two occasions (1st: one month before exam, 2nd*: on the first day of exam week)	Plasma IgA increased.
McClelland (1985)	Exam	+ (p<.06) (right after exam) +- (one and 3/4 hr later)	Three occasions (1st*: right after exam, 2nd*: one and 3/4 hr later, 3rd: several days later)	Rise of sIgA right after exam was followed by a drop one and 3/4 hr later. Lower sIgA, steeper drop of sIgA, and greater increase of NE in stronger n power group.
Evans (1994)	presentation for a science module	+(p<.1) (immediately after presentation)	In 2 consecutive weeks during the same scheduled hours (9: 30, 10: 30, 11: 30, 12: 30), and immediately after presentation.	Increase in cortisol.
Spangler (1997)	Exam	++ (p<.05) (5 and 15 min after exam)	15 min before and 5 and 15 min after exam	SigA also increased after the control situation.

*: stressful period RAS: the relaxed affiliative motive syndrome IPS: the inhibited power motive syndrome URT: upper respiratory tract

Possible mechanisms of variation of sIgA secretion caused by psychosocial factors.

As already shown in Fig.1, sIgA in saliva is produced locally by plasma cells in the salivary gland, and transported from the basolateral site of aciner cells or ductal cells of the salivary gland to their luminal site by the mechanism of SC dependent transport^{3,12}.

In the process of production and secretion of sIgA, we can point out three steps where psychosocial factors would affect a variation of sIgA secretion. In Fig.1, those steps are indicated by the large numerical characters, 1, 2, and 3.

The step shown by the character '1' in Fig.1 is the amount of plasma cells of the salivary gland. The number of plasma cells, expected to correlate positively with the level of IgA secretion in saliva, is in the balance with the continuous recruitment of plasma cells and the loss of them.

The plasma cells in the salivary gland will be recruited continuously to the gland in the following way. The precursor cells, B lymphocytes, are activated in mucosal associated lymphoid tissue (MALT), and induced switching to IgA. MALT includes gut associated lymphoid tissue (GALT), bronchial associated lymphoid tissue (BALT), and tonsil. The activated B lymphocytes migrate from MALT through lymphatic vessels into the blood circulation, then reach and stay in the salivary gland afterwards³⁷. Several days will pass before the B lymphocytes mature into plasma cells in the salivary gland.

At the same time, the plasma cells are lost continuously from the salivary gland, depending on the life span of the plasma cells. The average life span of plasma cells is thought to be less than 20 days, relating to the site of B cell activation, e.g. spleen, lymph nodes, lamina propria, and bone marrow³⁸.

Without the recruitment of precursor cells, continuous loss of plasma cells will result in a reduction of the number of plasma cells, and, consequently, in a decrease of sIgA secretion. In the salivary gland, rapid turnover of plasma cells was indicated in the case of chronic lymphocytic leukemia given an extracorporeal

blood irradiation therapy, and showing a significant decrease of circulating lymphocytes, which means a reduction in the supply of precursor cells of plasma cells, followed by a rapid decrease in salivary IgA levels³⁹.

Favorably, in meta-analytic studies, it is concluded that psychological stress affects T and B lymphocytes in the blood so as to inhibit the number and functions of the lymphocytes^{40,41}. Stress hormones, cortisol and catecholamines, may be responsible for this stress effect on lymphocytes⁴².

Since the effect of psychological stress on blood lymphocytes resembles that of extracorporeal blood irradiation, it is expected that psychological stress decreases salivary sIgA secretion in the same manner as extracorporeal blood irradiation, through a reduction in the recruitment of precursor cells of plasma cells to the salivary gland.

The decrease of sIgA secretion will happen gradually, depending on the reduction rate of the supplying precursor cells and the half life of plasma cells. It is tempting to think this speculated mechanism of a decrease in sIgA may underlie the delayed stress effect on the sIgA response, mentioned previously, meaning a decrease of sIgA secretion several days after stress.

Another step, shown by character '2' in Fig.1, is the IgA producing ability of plasma cells. There is no direct evidence that a psychological stress or relaxation modulates IgA production by plasma cells yet. However, psychological stress or relaxation may possibly modulate plasma cell activity through stimulation of nerves, cytokines, and hormones. This is considered because a histopathologic study showed that plasma cells in the lamina propria were associated with nerve fibers^{42,43}. Also, some studies reported that several cytokines, including interleukin-1 (IL-1), interleukin-3 (IL-3), interleukin-4 (IL-4), interleukin-5 (IL-5), interleukin-6 (IL-6), erythropoietin, and nerve growth factor^{44,51}, and hormones, including growth hormone, insulin-like growth factor-I, insulin-like growth factor-II, and insulin⁵², influence the immunoglobulin production by plasma cells.

The remaining step which psychosocial factors might affect, shown by character '3' in Fig.1, is the SC dependent transport of

Table 2 Effect of other psychosocial factors on salivary IgA²⁸⁻³¹.

Author	Factor	Change or Relation	Sampling time	Note
Farne (1992)	Variation of daily hassles	--(p<.01) (4 weeks after)	Three times with a 4 week interval.	
Evans (1993)	Negative mood	Positive correlation (r= .39, p<.006)	Each evening up to 2 weeks	Between subjects, 'net' desirable events positively correlated with sIgA (r= .56, p< .028).
Stone (1994)	Desirable daily events	Positive correlation (r= .11, p<.01)	At the end of each day, for 12 weeks	Secretory IgA antibody activity to rabbit albumin was assayed.
	Undesirable daily events	Negative correlation (r= -.17, p<.05)		
Zeier (1996)	Work of air traffic controllers	++ (p<.001)	Before and after 100 min working session	Increase in salivary cortisol concentration (p<.001)

IgA by epithelial cells. The production of a secretory component by epithelial cells can be regulated by various kinds of immunological, endocrinary, and neural factors. These factors include interferon- γ ^{54,55}, IL-4⁵⁵, tumor necrosis factor- α ^{56,57}, IL-1- α ^{56,57}, IL-1- β ⁵⁸, estradiol⁵⁹ and androgens⁶⁰, cholinergic agonist^{57,58} and beta-adrenergic agonist^{57,58}, prostaglandin E2^{57,58}, vasoactive intestinal peptide^{57,58}, and somatostatin⁵⁸. However, it should be noted that the mode of effect of those factors is rather organ specific⁵⁸. Cholera toxin, 8-bromoadenosine 3':5'-cyclic monophosphate (bcAMP), and 8-bromoguanosine 3':5'-cyclic monophosphate (bcGMP) significantly increased the secretory component production by submandibular acinar cells^{57,58}. This result implies that, in the salivary gland, sIgA transport by the SC dependent system can be affected by psychological stress or relaxation through neural and/or endocrine mechanisms which vary intracellular cAMP or cGMP.

Can sIgA be a useful immunological stress marker?

As shown previously, current literature showed a favorable (positive) agreement in the effect of relaxing factors, whereas it showed a disagreement in the effect of stress on the sIgA response.

Stone et al⁶¹ attributed the inconsistency of the sIgA response

to a negative correlation between sIgA concentration and salivary flow⁶² and to a possible degradation of sIgA by proteases in whole saliva. Moreover, they proposed their original method of measuring specific sIgA antibodies in parotid saliva as a measure of immunocompetence, in place of measuring total sIgA in whole saliva⁶¹.

In opposition to the opinion of Stone et al⁶¹, Jemmott et al¹⁷ suggested that the negative correlation between the sIgA concentration and salivary flow is weak and not so problematic when unstimulated saliva is used, and taking salivary flow into account when measuring sIgA levels, e.g. assessing sIgA secretion rate, the problem of the negative correlation between sIgA concentration and salivary flow will be solved. Jemmott et al¹⁷ also suggested that the measurement of sIgA concentration in whole saliva is highly reproducible and stable over time. Finally, they concluded that sIgA is a useful measure of resistance to infectious diseases¹⁷.

It is true that Jemmott et al^{20,21} have successfully showed that academic stress inhibits the sIgA secretion rate. In addition, they showed personality characteristics differentiated patterns of sIgA secretion rate²⁰ and a positive effect of social support on sIgA levels²¹. (Refer notes in Table 1.)

However, Mouton et al²² suggested that assaying salivary IgA to measure stress may not be as useful in psychophysiological

Table 3 Effect of relaxing factors on salivary IgA³²⁻³⁶.

Author	Factor	Change	Sampling time	Note
Dillon (1985)	Humorous videotape	++ (p=.026)	Before and after videotape (30 min long)	
	Didactic videotape	+-		
Green (1987)	Relaxation response	++ (p<.05)	Before and after treatment (20 min long)	No significant change in salivary cortisol.
	Guided visualization	++ (p<.05)		
	Massage	++ (p<.01)		
	Lying down	+-		
	Touching control	+-		
Jasnoski (1987)	Relaxation	++ (p<.025) (vs vigilance task control)	Before and after protocol (1 hr long)	Negative correlation (r=-.39; p<.05) between sIgA and saliva norepinephrine.
Green (1988)	Relaxation	++ (p<.001)	Before and after 20-min relaxation practice	Increase of serum IgA (p<.001), IgG (p<.001), IgM (p<.05) over a 3 week practice period.
		++ (p=.014) (22nd day vs 1st day)	On the 22nd day of relaxation practice	
Olness (1989)	Self-hypnosis	+-	Before and after self-hypnosis (25 min long)	Children (6-12 yr old) were recruited.
	Self-hypnosis + specific suggestions	++ (p=.007)		

research as expected. It was because, in their academic stress study enrolling dentistry students on four occasions over a period of 8 months, they observed a significant difference in the level of salivary IgA only for the most polarized contrast, i.e., between final exam and end of summer vacation, and they also observed a weak negative correlation between the level of salivary IgA and the stress rating only at the final exam²².

In our opinion, the sIgA response to psychological stress can be rather complex because of the following. The delayed stress effect, which decreases sIgA secretion in saliva gradually, must be superimposed with the immediate stress effect, which increases sIgA secretion in saliva immediately. Thus, an inhibition of the sIgA response caused by stress experienced several days previously can be cancelled at the time of saliva sampling by an increase in the sIgA response caused by another stress experienced only several hours before saliva sampling.

Therefore, when the delayed and inhibitory effect of some stress on the sIgA response is concerned, it is necessary to remove immediate and increasing effect of other stresses from the sIgA response. Deinzer, et al.²³ successfully solved the problem by sampling saliva every morning immediately after awakening and before doing anything else.

Reversely, when the immediate and increasing effects of some stress on the sIgA response is concerned, the delayed and the inhibitory effect of other stresses would not affect the result, because the delayed stress effect would be almost constant during the short experimental period. From this point, sIgA in saliva can

be said to be a more suitable immunological marker for the immediate stress effect than for the delayed stress effect.

Complexity of sIgA response to stress may be seen only in human. Interestingly, it is reported that salivary IgA can be a marker of social stress in rats⁶³. Male rats housed singly showed stable sIgA levels with little variation, while those housed separately with a female showed an initial decrease in sIgA followed by a steady increase. Males housed in a group (n=6) showed a steady decline in sIgA levels⁶³.

In their study of air traffic controllers, Zeier, et al.³¹ suggested that positive emotional engagement is responsible for the observed sIgA increase. They also suggested that measuring sIgA response may be a valuable tool for differentiating between positive and negative stress effects or between successful and unsuccessful adaptation or coping with situational demands³¹.

Miletic, et al.⁶⁴ reported an interesting fact that elderly persons who were excited by social events over the weekend days (friends or family member visits, worship and social events in churches, etc.) showed an increase in their sIgA secretion during the weekend days.

In summary, sIgA in saliva is a promising candidate as a stress marker which may be able to differentiate between positive and negative stress effects. However, some additional studies will be necessary to establish proper methods to separate the immediate stress effect and the delayed stress effect on sIgA response, and to elucidate mechanisms of sIgA variation caused by psychosocial factors.

References

- 1) Jemmott JB III, Locke SE. Psychosocial factors, immunologic mediation, and human susceptibility to infectious diseases: How much do we know? *Psychol Bull* 1984; 95: 78-108.
- 2) Kiecolt-Glaser JK, Glase R. Stress and immune function in humans. In: Ader R, Felton D, Cohen E., eds. *Psychoneuroimmunology*, 2nd ed. Orlando: Academic Press, Inc., 1991: 849-67.
- 3) Tomasi TB. Structure and function of mucosal antibodies. *Annu Rev Med* 1970; 21: 281-98.
- 4) Hanson LA, Ahlstedt S, Andersson B, et al. Mucosal immunity. *Ann N Y Acad Sci* 1983; 409: 1-21.
- 5) Mestecky J, Russell MW, Jackson S, Brown TA. The human IgA system: A reassessment. *Clin Immunol Immunopathol* 1986; 40: 105-14.
- 6) Brandtzaeg P, Bjerke K, Kett K, et al. Production and secretion of immunoglobulins in the gastrointestinal tract. *Ann Allergy* 1987; 59: 21-39.
- 7) Mestecky J, Czerkinsky C, Russell MW, et al. Induction and molecular properties of secretory and serum IgA antibodies specific for environmental antigens. *Ann Allergy* 1987; 59: 54-9.
- 8) Mestecky J, McGhee JR. Immunoglobulin A (IgA): Molecular and cellular interactions involved in IgA biosynthesis and immune response. *Adv Immunol* 1987; 40:153-245.
- 9) Childers NK, Bruce MG, McGhee JR. Molecular mechanisms of immunoglobulin A defense. *Annu Rev Microbiol* 1989; 43: 503-36.
- 10) Aroeti B, Casanova J, Okamoto C, et al. Polymeric immunoglobulin receptor. *Int Rev Cytol* 1992; 137: 157-68.
- 11) Mostov KE. Transepithelial transport of immunoglobulins. *Annu Rev Immunol* 1994; 12: 63-84.
- 12) Tomasi TB. Introduction:an overview of the mucosal system. In: Orga PL, Strober W, Mestecky J, McGhee JR, Lamm ME, Bienenstock J, eds. *Handbook of mucosal immunology*. San Diego: Academic Press, Inc., 1994: 3-8.
- 13) Armstrong SJ, Dimmock NJ. Neutralization of influenza virus by low concentrations of hemagglutinin-specific polymeric immunoglobulin A inhibits viral fusion activity, but activation of the ribonucleoprotein is also inhibited. *J Virol* 1992; 66: 3823-32.
- 14) Mazanec MB, Kaetzel CS, Lamm ME, Fletcher D, Nedrud JG. Intracellular neutralization of virus by immunoglobulin A antibodies. *Proc Natl Acad Sci USA* 1992; 89: 6901-5.
- 15) Silbart LK, Keren DF. Reduction of intestinal carcinogen absorption by carcinogen-specific secretory immunity. *Science* 1989; 243: 1462-4.
- 16) Kaetzel CS, Robinson JK, Chintalacharuvu KR, Vaerman J-P, Lamm ME. The polymeric immunoglobulin receptor (secretory component) mediates transport of immune complexes across epithelial cells: A local defense function for IgA. *Proc Natl Acad Sci USA* 1991; 88: 8796-800.
- 17) Jemmott JB, McClelland DC. Secretory IgA as a measure of resistance to infectious disease: comments of Stone, Cox, Valdimarsdottir, and Neale. *Behav Med* 1989; 15: 63-71.
- 18) Gleeson M, Cripps AW, Clancy RL, Hensley MJ, Henry RJ, Wlodarczyk JH. The significance of transient mucosal IgA deficiency on the development of asthma and atopy in children. In: Mestecky J, et al. eds. *Advances in Mucosal*

- Immunology. New York: Plenum Press, 1995:861-4.
- 19) Drummond PD, Hewson-Bower B. Increased psychosocial stress and decreased mucosal immunity in children with recurrent upper respiratory tract infection. *J Psychosom Res* 1997; 43 (3): 271-8.
 - 20) Jemmott JB III, Borysenko JZ, Borysenko M, et al. Academic stress, power motivation, and decrease in secretion rate of salivary secretory immunoglobulin A. *Lancet* 1983; 1: 1400-2.
 - 21) Jemmott JB III, Magloire K. Academic stress, social support, and secretory immunoglobulin A. *J Pers Soc Psychol* 1988; 55 (5): 803-10.
 - 22) Mouton C, Fillion L, Tawadros E, Tessier R. Salivary IgA is a weak stress marker. *Behav Med* 1989; 15 (4): 179-85.
 - 23) Deinzer R, Schuller N. Dynamics of stress-related decrease of salivary immunoglobulin A (sIgA): Relationship to symptoms of the common cold and studying behavior. *Behav Med* 1998; 23: 161-9.
 - 24) Kiecolt-Glaser JK, Garner W, Speicher C, Penn GM, Holliday J, Glaser R. Psychosocial modifiers of immunocompetence in medical students. *Psychosom Med* 1984; 46 (1): 7-14.
 - 25) McClelland DC, Ross G, Patel V. The effect of an academic examination on salivary norepinephrine and immunoglobulin levels. *J Human Stress* 1985; 11: 52-9.
 - 26) Evans P, Bristow M, Hucklebridge F, Clow A, Pang F-Y. Stress, arousal, cortisol and secretory immunoglobulin A in students undergoing assessment. *Br J Clin Psychol* 1994; 33: 575-6.
 - 27) Spangler G. Psychological and physiological responses during an exam and their relation to personality characteristics. *Psychoneuroendocrinology* 1997; 22 (6): 423-41.
 - 28) Farne M, Boni P, Gnugnoli D, Corallo A. The effect of daily hassles on salivary IgA: experimental evidence. *Boll Soc Ital Biol Sper* 1992; 68: 409-12.
 - 29) Evans P, Bristow M, Hucklebridge F, Clow A, Walters N. The relationship between secretory immunity, mood, and life events. *Br J Clin Psychol* 1993; 32: 227-36.
 - 30) Stone AA, Neale JM, Cox DS, Napoli A, Vldimarsdottir H, Kennedy-Moore E. Daily events are associated with a secretory immune response to an oral antigen in men. *Health Psychol* 1994; 13: 440-6.
 - 31) Zeier H, Brauchli P, Joller-Jemelka HI. Effects of work demands on immunoglobulin A and cortisol in air traffic controllers. *Biol Psychol* 1996; 42: 413-23.
 - 32) Dillon KM, and Minchoff B. Positive emotional states and enhancement of the immune system. *Int J Psychiatry Med* 1985-86; 15 (1): 13-8.
 - 33) Green RG, Green M. Relaxation increases salivary immunoglobulin A. *Psychol Rep* 1987; 61: 623-9.
 - 34) Jasnoski ML, Kugler J. Relaxation, imagery, and neuroimmuno-modulation. *Ann N Y Acad Sci* 1987;496: 722-30.
 - 35) Green ML, Green RG, Santoro W. Daily relaxation modifies serum and salivary immunoglobulins and psychophysiologic symptom severity. *Biofeedback Self Regul* 1988; 13 (3): 187-99.
 - 36) Olness K, Culbert T, Uden D. Self-regulation of salivary immunoglobulin A by children. *Pediatrics* 1989; 83 (1): 66-71.
 - 37) Croitoru K, Bienenstock J. Characteristics and functions of mucosa-associated lymphoid tissue. In: Orga PL, Strober W, Mestecky J, McGhee JR, Lamm ME, Bienenstock J, eds. *Handbook of mucosal immunology*. San Diego: Academic Press, Inc., 1994: 141-9.
 - 38) MacLennan ICM, Gray D. Antigen-driven selection of virgin and memory B cells. *Immunol Rev* 1986; No91: 61-85.
 - 39) Friedman BK, Greenberg B. Effect of extracorporeal cesium irradiation on secretory and serum immunoglobulins. *J Dent Res* 1975; 54; special issue A: 68.
 - 40) Herbert TB, Cohen S. Stress and immunity in humans: a meta-analytic review. *Psychosom Med* 1993; 55: 364-79.
 - 41) van Rood YR, Bogaards M, Goulmy E, van Houwelingen HC. The effects of stress and relaxation on the in vitro immune response in man: a meta-analytic study. *J Behav Med* 1993; 16: 163-81.
 - 42) Stratakis CA, Chrousos GP. Neuroendocrinology and pathophysiology of the stress system. *Ann N Y Acad Sci* 1995; 771: 1-18.
 - 43) Cauna N, Cauna D. Association of nerve fibers and plasma cells in abnormal human nasal respiratory mucosa. *Ann Otol Rhinol Laryngol* 1974; 83: 347-59.
 - 44) Stead RH. Innervation of mucosal immune cells in the gastrointestinal tract. *Reg Immunol* 1992; 4 (2): 91-9.
 - 45) Kawano M, Tanaka H, Ishikawa M, et al. Interleukin-1 accelerates autocrine growth of myeloma cells through interleukin-6 in human myeloma. *Blood* 1989; 73: 2145-8.
 - 46) Bergui L, Schena M, Gaidano G, Riva M, Caligaris-Cappio F. Interleukin 3 and interleukin 6 synergistically promote the proliferation and differentiation of malignant plasma cell precursors in murine multiple myeloma. *J Exp Med* 1989; 170: 613-8.
 - 47) Anderson KC, Jones RM, Morimoto C, Leavitt P, Barut BA. Response patterns of purified myeloma cells to hematopoietic growth factors. *Blood* 1989; 73: 1915-24.
 - 48) Taylor CW, Grogan TM, Salmon SE. Effects of interleukin-4 on the in vitro growth of human lymphoid and plasma cell neoplasms. *Blood* 1990; 75: 1114-8.
 - 49) Kawano M, Hirano T, Matusda T, et al. Autocrine generation and requirement of BSF-2/IL-6 for human multiple myelomas. *Nature* 1988; 332: 83-5.
 - 50) Kimata H, Sherr EH, Saxon A. Human natural killer (NK) cells produce a late-acting B-cell differentiation activity. *J Clin Immunol* 1988; 8: 381-9.
 - 51) Kimata H, Yoshida A, Ishioka C, Mikawa H. Erythropoietin enhances immunoglobulin production and proliferation by human plasma cells in a serum-free medium. *Clin Immunol Immunopathol* 1991; 59: 495-501.
 - 52) Kimata H, Yoshida A, Ishioka C, Mikawa H. Nerve growth factor inhibits immunoglobulin production by but not proliferation of human plasma cell lines. *Clin Immunol Immunopathol* 1991; 60: 145-51.
 - 53) Kimata H, Yoshida A. Differential effect of growth hormone and insulin-like growth factor-I, insulin-like growth factor-II, and insulin on Ig production and growth in human plasma cells. *Blood* 1994; 83 (6): 1569-74.
 - 54) Sollid LM, Kvale D, Brandtzaeg P, Markussen G, Thorsby E. Interferon-gamma enhances expression of secretory component, the epithelial receptor for polymeric immunoglobulins. *J Immunol* 1987; 138: 4303-6.
 - 55) Phillips JO, Everson MP, Moldoveanu Z, Lue C, Mestecky

- J. Synergistic effect of IL-4 and IFN-gamma on the expression of polymeric Ig receptor (secretory component) and IgA binding by human epithelial cells. *J Immunol* 1990; 145: 1740-4.
- 56) Kvale D, Lovhaug D, Sollid LM, Brandtzaeg P. Tumor necrosis factor-alpha up-regulates expression of secretory component, the epithelial receptor for polymeric Ig. *J Immunol* 1988; 140: 3086-9.
- 57) Kelleher RS, Hann LE, Edwards JA, Sullivan DA. Endocrine, neural, and immune control of secretory component output by lacrimal gland acinar cells. *J Immunol* 1991; 146 (10): 3405-12.
- 58) Lambert RW, Gao J, Kelleher RS, Wickman LA, Sullivan DA. Neural, endocrine and immune regulation of secretory component production by lacrimal gland acinar cells. In: Mestecky J, et al. eds. *Advances in Mucosal Immunology*. New York: Plenum Press, 1995: 221-4.
- 59) Sullivan DA, Wira CR. Variations in free secretory component levels in mucosal secretions of the rat. *J Immunol* 1983; 130 (3): 1330-5.
- 60) Sullivan DA, Kelleher RS, Vaerman J-P, Hann LE. Androgen regulation of secretory component synthesis by lacrimal gland acinar cells in vitro. *J Immunol* 1990; 145 (12): 4238-44.
- 61) Stone AA, Cox DS, Valdimarsdottir H, Neale JM. Secretory IgA as a measure of immunocompetence. *J Human Stress* 1987; 13: 136-40.
- 62) Brandtzaeg P. Human secretory immunoglobulins VII, concentrations of parotid IgA and other secretory proteins in relation to the rate of flow and duration of secretory stimulus. *Arch Oral Biol* 1971; 16: 1295-310.
- 63) Guhad FA, Hau J. Salivary IgA as a marker of social stress in rats. *Neurosci Lett* 1996; 216: 137-40.
- 64) Miletic ID, Schiffman SS, Miletic VD, Sattely-Miller EA. Salivary IgA secretion rate in young and elderly persons. *Physiol Behav* 1996; 60 (1): 243-8.